A TECHNIQUE TO MEASURE STRAIN DISTRIBUTIONS IN SINGLE WOOD PULP FIBERS

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ABSTRACT

Environmental scanning electron microscopy (ESEM) and digital image correlation (DIC) were used to measure microstrain distributions on the surface of wood pulp fibers. A loading stage incorporating a fiber gripping system was designed and built by the authors. Fitted to the tensile stage of an ESEM or a Polymer Laboratories MINIMAT tester, it provided a reliable fiber straining mechanism. Black spruce latewood fibers (Picea mariana (Mill) B.S.F.) of a near-zero microfibril angle displayed a characteristically linear load-elongation form. ESEM was able to provide real-time, high magnification images of straining fibers, crack growth, and complex single fiber failure mechanisms. Digital images of single fibers were also captured and used for subsequent DIC-based strain analysis. Surface displacement and strain maps revealed nonuniform strain distributions in seemingly defect-free fiber regions. Applied tensile displacements resulted in a strain band phenomenon. Peak strain (concentration) values within the bands ranged from 0.9% to 8.8%. It is hypothesized that this common pattern is due to a combination of factors including the action of microcompressive defects and straining of amorphous cell-wall polymeric components. Strain concentrations also corresponded well to locations of obvious strain risers such as visible cell-wall defects. Results suggest that the ESEM-based DIC system is a useful and accurate method to assess and, for the first time, measure fiber micromechanical properties.

Keywords: Fibers, micromechanics, strain, digital image correlation, tensile testing, environmental scanning electron microscopy.

INTRODUCTION

Single fiber tensile tests conducted under polarized light confirmed that the microfibril angle of the S2 secondary cell-wall layer largely controls the mechanical properties of individual wood pulp fibers (Page et al. 1977). By the same technique, fiber defects have also been shown to influence fiber properties including maximum strain potential and fiber failure.
mechanisms. However, progress with regard to characterizing mechanical properties of recycled fibers (Wuu et al. 1991) and other fiber characteristics, such as fracture criteria, is slow; and no empirical data exist that can quantify such basic relationships as (collapsed) fiber cell-wall Poisson ratios. Evidence of individual defect impact on the mechanical response of the cell wall is lacking, and at present statistical descriptions of fiber defect/length relationships represent the extent of fiber micromechanics understanding (Page and El-Hosseiny 1976). The objective of this study was to develop a single fiber tensile testing technique capable of extracting micro mechanical property data from reliable single fiber tensile tests. Of particular interest was the ability to observe fracture sequences in single fibers and measure microstrain distributions surrounding natural and processing-induced defects.

MATERIALS AND METHODOLOGY

Single fibers were observed under load using environmental scanning electron microscopy (ESEM). The ESEM is similar to conventional scanning electron microscopes (SEM) in that high magnification and large depth of field images can provide insight into material characteristics. However, the unique design of the ESEM column and secondary electron detector permit the specimen chamber to operate under increased pressures of 5–20 Torr (Cameron and Donald 1994). Water vapor is often used to provide the chamber atmosphere, in which case it is possible to image biological specimens without subjecting them to potentially harmful dehydration processes or conductive coating. A chamber water vapor pressure of approximately 6 Torr was used in this study. Under such conditions single wood pulp fibers were examined in the ESEM in their near natural state. Additional information on this instrument is available in the literature (Cameron and Donald 1994; Sheehan and Scriven 1991).

ESEM observation of single fiber testing

In an effort to establish the reliability of mechanical property data obtained from the developed free-fiber-alignment gripping mechanism, various single fiber types were tested using the loading stage in combination with the ESEM and a Polymer Laboratories MINI-MAT tensile tester. The fiber gripping system was based upon a ball and socket mechanism (Fig. 1). Epoxy droplets applied close to the fiber ends prior to testing provided the ball joints. Although this assembly has been proven to reduce fiber failure at the grips by up to 40% (Mott 1995), it does not completely remove the influence that gripping mechanisms have upon stress and strain distributions within the fiber. For the purposes of this study, however, surface strain was most frequently measured close to the fiber midspan, and a common fiber length to width ratio was approximately 30:1. For such geometries, even in highly anisotropic plates where the average fibril angle approaches 15°, the effect of gripping on strain component distribution at midspan is negligible (Wu and Thomas 1968). The effect is further reduced in near-zero microfibril angle fibers because the collapsed fiber is not a single orthotropic plate. Instead, the opposite walls of the fiber restrict the shear deformation that results from gripping, which
frequently leads to normal stress-strain non-uniformity.

More details on this technique and preliminary findings of this study, which include ESEM failure and crack initiation sequences and a critical assessment of fiber macromechanical property data, are reported elsewhere (Mott 1995; Mott et al. 1995; Groom et al. 1995). This paper describes a micromechanical investigation of fiber cell-wall material using digital image correlation.

Single fibers were introduced into the ESEM specimen chamber under an ambient equilibrium moisture content. A carefully controlled (vacuum) pump-down schedule for the ESEM chamber was used to maintain fibers in their natural hydrated state (Cameron and Donald 1994). The specimen chamber pump-down sequence was characterized by an 8-stage water vapor flooding cycle while simultaneously positioning the fiber out of the beam path to reduce the potential for radiation damage. The final ESEM operating parameters for observing single fibers were a chamber pressure of 5 to 6.3 Torr and a substage temperature of approximately 21°C. Under these carefully monitored environmental conditions, a single fiber equilibrium moisture content of approximately 8.5% was calculated using the Carrier equation (Siau 1984).

The wet bulb depression temperature (a required variable in the Carrier equation) under established ESEM test conditions was determined by first obtaining the corresponding dew point temperature using a Peltier cooling substage. Corresponding dry bulb and the dew point temperatures were then used to find the equivalent wet bulb depression temperature (12°C) with the aid of a psychrometric chart (Siau 1984). A condenser lens strength of 56-60% (instrument setting) and an accelerating voltage of 15 keV were used when examining the fibers.

Digitally captured ESEM images for DIC

For surface-microstrain assessment purposes, single black spruce (Picea mariana (Mill) B.S.P.) latewood fibers were obtained by batch kraft pulping the 55th growth ring of a mature tree to a 47% yield (kappa number 56). During preparation, single fibers were also flattened and dried into ribbon shapes prior to tensile testing. More information concerned with fiber drying, moisture content equilibration, and adhesive application is reported in Mott (1995) and Mott et al. (1995). Fiber drying and flattening facilitate easy handling and permit treatment of the fiber as an orthotropic bilaminate material (Page et al. 1977). Flattened fibers also simulated collapsed single fiber geometries in paper sheets and other biocomposites.

Digitally captured (640 x 480 pixel) images of fibers under a uniaxial tensile displacement were used to assess surface microstrains. The experimental setup is presented in Fig. 1. Obtaining suitably high signal to noise ratio images of straining fibers required that a slow electron beam scan rate of 4.3 s/frame be employed. This precluded any further need for image averaging, which is often necessary when employing DIC analysis (James et al. 1989). Following completion of the pump-down and flooding procedure, single fibers were assumed to be under the calculated equilibrium conditions (this process can take up to 2 min). The tensile substage was then moved into the beam path. A low magnification (×300) and an increased scan rate (2.3 frames/s) were selected together with an increased working distance of approximately 10 mm. This procedure was adopted to reduce the potentially damaging effects of the electron beam. Despite visible image noise, fiber surface features such as pit fields were plainly evident, and a fiber gauge length was accurately measured using resident ESEM software (±1 μm).

A suitable area over which strain analysis was to be performed was quickly identified, and the necessary ESEM parameters were adjusted to obtain optimum image quality. A final magnification of approximately 2,500 was usually selected prior to commencing tensile displacement at a rate of 1 μm/s (console controlled). Optimum brightness, contrast levels, beam and condenser strengths were achieved
Measurement of the microstrain field distributions using DIC required obtaining two successive digital ESEM images over an approximate 5-s displacement (or loading) interval. These images were designated an image pair (Fig. 2). Prior to obtaining any images for DIC, each fiber was loaded so as to achieve a linear load elongation trace. Extension was then suspended and an initial image captured to a remote PC resident framegrabber (approximately 1/30 s capture time). Suspending the tensile test is essential when employing slow scan rates in dynamic microscopy. Failure to do so results in time-delay distorted images where pixels in the lower portion of an image are temporally distorted due to the fact that they are representative of a specimen that has been subject to a proportionally greater strain.

After capturing the initial image, displacement was resumed employing a cross-head extension rate of 1 μm/s for approximately 5 s. Prior to capturing a second image, extension was again suspended. The acquired image pair was then used to perform a DIC analysis of fiber surface displacements resulting from both the 5-μm fiber extension and the small irregular (purely) translational movements of the tensile substage.

Following image acquisition, each fiber was also strained to failure in the ESEM. This required subjecting each fiber to approximately 15–20 s more beam time. Fracture surfaces were then used to assess whether beam damage had significantly affected mechanical properties. The black spruce fibers used in this study demonstrated a brash fracture surface, as is common with mature fibers of low microfibril angle (Fig. 3). The fibers were also prone to cracking and failure at locations other than regions examined by ESEM. These facts provided an initial confirmation that beam-related radiation damage was not significantly affecting the mechanical response of the fiber cell-wall material. Average strain to failure for 218 black spruce latewood fibers as determined by the MINIMAT testing facility was approximately 4.25%.

**Principles of DIC**

Surface displacements of fibers were determined using a digital image correlation (DIC) technique. Digital image pairs were analyzed using a DIC program written to utilize a Silicon Graphics workstation (Mott 1995). Bi-cubic splines were used to generate continuous surface intensity fields from the discrete digital images of the tensile substage. The processes described above were performed for each fiber, resulting in a set of data that could be used to study the mechanical properties of fibers under different conditions.
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FIG. 4. A digitally captured image of a single wood fiber. The fiber was slowly dried between glass slides to exploit the phenomenon of anisotropic shrinkage. The resulting shrinkage patterns created sufficient image contrast to permit subsequent DIC assessment of surface displacements.

DIC accuracy and calibration

To obtain the necessary sub-pixel accuracy, a continuous intensity function bi-cubic interpolation routine was applied to respective ESEM image pairs. This made it feasible to measure intensity values between discrete pixel locations and to measure respective displacements with a sub-pixel precision of ± 0.1 pixel. At a magnification of 2,500 this was equal to approximately ± 0.008 μm. This value was determined by a rigid body translation technique, conducted digitally and then mechanically inside the ESEM chamber.
Digital warping of a single duplicated image was performed by means of a Silicon Graphics library routine. This provided evidence of the DIC program's ability to determine displacements in a rapid and efficient fashion. Applied strains in the x,y plane were recovered with no reported error. This was to be expected in such noise-free images. True noise-influenced (displacement) measurement precision was established using mechanically translated ESEM images. Translation was conducted by moving the tensile substage small amounts in the x,y plane. Mean tolerances in the x and y axes were then determined by running 19 sets of correlation arrays per image pair (4 image pairs). Each set consisted of a regular 14 x 14 pixel array (Fig. 5). Analyses of variances (ANOVA) and subsequent multiple comparison of means tests (Tukey's multiple range test) indicated that no statistically significant differences existed between the reported mean values for u and v displacement between any of the chosen 19 data sets ($P = 0.00$). It was concluded, therefore, that the system was not affected by spatially nonrandom noise. This was confirmed in subsequent tests using the same 19 data set locations applied to translations of varying magnitude. Mean u and v standard deviation were reported as tolerance value of 0.08 pixels and 0.1 pixels, respectively.

To visualize single fiber surface displacement due to microextensometry-induced deformation, the u and v (horizontal and vertical) components of displacement were plotted as 3-D surface maps. Examples of u and v displacement component vectors for a black spruce late-wood fiber are presented in Fig. 6. The u-component of displacement represents passive deformation in the fiber transverse axis resulting from an applied tensile displacement in the fiber longitudinal axis (v). Displacement vector v represents the longitudinal response of the cell wall to same applied tensile displacement.

**Determination of strains**

Surface strain values were determined by first calculating the change in distance (in pixel units) between respective neighboring subre-
region locations resulting from tensile displacement. The Lagrangian strain tensor definition was then used to obtain normal strains in the x and y directions. The system dependent sub-pixel precision level (± 0.1 pixel) required that the control points of subregions have a 20 pixel spacing in order to obtain a strain measurement precision of ± 0.5% or better. To obtain a strain precision of ±1% required a subimage spacing of only 10 pixels (0.77 µm at a 2,500 magnification).

Figure 7 displays rendered strain maps for single black spruce latwood fiber. Accompanying displacement surfaces are displayed in Fig. 6. The fiber contained a number of microcompressive kinks or creases resulting from a modified preparation technique whereby two fibers were dried, one bridging the other. Strain components εxx and εyy indicated that a creased fiber cell wall results in large strain variations. A rational explanation of strain nonuniformity is offered.

**Fiber strain distributions**

The ESEM-based DIC system was successfully used for calculating the strain distributions of six defect-free black spruce latwood fibers under small applied strains of 0.3% to 0.5%. Component εyy indicated that straining in the direction of loading is distinctly non-uniform. Banded regions of high strain concentrations (0.9% to 8.8%), similar in character to Luders' bands which form in some metals (Illston et al. 1987), developed in the fiber wall under uniaxial tensile displacements. εxx was more uniform although strain concentrations of a lesser magnitude were evident. εxy was also largely negative, providing confirmatory evidence of a Poisson effect. These patterns were common to all defect-free fibers of black spruce latwood with a low microfibril angle tested in this preliminary study, the results of which are discussed in more detail in other sources (Mott 1995; Mott et al. 1995; Groom et al. 1995).

A reasonable explanation for strain non-uniformity was thought to be the manifestation of microcompressive defect "pull-out."

Microcompressive pull-out occurs as small crimps, creases, and kinks are removed from the cell wall under increasing tensile strain. Evidence of the high strain potential in microcompressively damaged regions is also
found when examining a fiber load elongation curve, which is frequently irregular and shallow in initial stages as the integrated effects of microcompressive pull-out take effect (Page and Seth 1980). A further potential cause of the strain band effect in the yy axis that cannot be discounted is the periodic and natural occurrence of amorphous cellulose regions and the presence of interstitial lignin or hemicellulose polymer phases of the cell-wall composite structure.

An initial assumption of DIC theory is that strain within any chosen subregion is uniform (Chu et al. 1985). This assumption was deemed valid when testing defect-free and minimally defected fibers. However, highly localized plastic deformations, such as those which occur around the tip of a growing crack in ductile materials, are problematic and would be difficult to measure unless small enough subregion areas were attainable. ESEM video microextensometry indicated that single fibers frequently display what can be considered a ductile cracking mechanism, characterized by a blunt crack tip. These cracks are commonly seen to extend from pit apertures and pit borders under increasing tensile displacement. Strain map pairs were rendered for several fibers that included a visible bordered pit. High strain concentrations were found to correspond with common displacement or load-induced crack locations at the pit aperture and at the pit border where the greatest S2 layer microfibrillar deviation is known to occur.

The strain values close to the pit aperture were extremely large, however, and although representative of regions where high strain concentrations are likely to occur (they correspond to actual crack locations), the values are likely in error. The probable source of this error is a violation of the DIC assumption that strains within subregions be uniform (Chu et al. 1985). The failure of DIC to predict strain values in the locality of pits supported the hypothesis that large strain gradients exist in small regions (< 1 μm²) immediately adjacent to the aperture edge and that highly localized plastic deformation of this region was probable. A confirmed premature yielding of this type would have serious implications with regard to damage accumulation in single fibers subject to low repetitive or cyclic loading. The probable and potentially catastrophic premature plastic deformation of the cell wall in this study occurred in single fibers that were subject to applied strains of 1% or less.

CONCLUSIONS

ESEM-based DIC was used to determine single fiber microstrain distributions. Using present ESEM and DIC capabilities and carefully controlled fiber preparation techniques, it has been possible for the first time to measure single wood fiber Poisson effects, to quantify natural and induced cell-wall defects in terms of strain potential, and to explore and quantify single fiber failure mechanisms. Expected strain nonuniformity has been measured, and evidence for a quasi-ductile fracture mechanism and of a premature plastic failure region in the locality of natural defects now exists. For such analyses, a ±0.1 pixel displacement measurement error and an optimum DIC subimage area of 29² pixels was adequate.

Smaller DIC measurement error, in the order of 0.01 pixel, now appears feasible using improved DIC algorithms (Lif et al. 1995). Improved computing facilities used in combination with such algorithms will promote faster DIC analyses and more precise data to confirm the initial findings of this study. For the present, however, ESEM-based DIC provides a powerful new tool to investigate the micromechanical process of fiber deformation and fracture, which together provide the bridge between fiber composite structure and morphology and mechanical properties. Investigation of this process is fundamental to the improvement of biocomposite materials.

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