

## Research and Education

**Cite this article:** Enloe SF, Lucardi RD, Loewenstein NJ, Lauer DK (2018) Response of twelve Florida cogongrass (*Imperata cylindrica*) populations to herbicide treatment. *Invasive Plant Sci Manag* 11:82–88. doi: 10.1017/inp.2018.12

Received: 5 September 2017

Revised: 28 March 2018

Accepted: 3 April 2018

### Associate Editor:

James K. Leary, University of Hawaii at Manoa

### Key words:

Invasive grass control; herbicide antagonism

### Author for correspondence:

Stephen F. Enloe, Department of Agronomy, University of Florida, Gainesville, FL 32653. (Email: sfenloe@ufl.edu)

# Response of twelve Florida cogongrass (*Imperata cylindrica*) populations to herbicide treatment

Stephen F. Enloe<sup>1</sup>, Rima D. Lucardi<sup>2</sup>, Nancy J. Loewenstein<sup>3</sup> and Dwight K. Lauer<sup>4</sup>

<sup>1</sup>Associate Professor, Department of Agronomy, University of Florida, Gainesville, FL, U.S.A., <sup>2</sup>Research Ecologist, Forestry Science Laboratory, USDA Forest Service Southern Research Station, Athens, GA, U.S.A., <sup>3</sup>Extension Specialist, School of Forestry and Wildlife Sciences, Auburn University, Auburn, AL, U.S.A. and <sup>4</sup>Analyst, Silvics Analytic, Wingate, NC, U.S.A.

## Abstract

Cogongrass [*Imperata cylindrica* (L.) Beauv.] is an invasive grass in the southeastern United States, and its impacts strongly affect the region, especially Florida. Herbicide strategies have been limited to glyphosate in natural areas and imazapyr in managed pine forests and non-crop areas where its soil residual activity is of less concern. This lack of options has raised concern for herbicide resistance, which has never been documented for *I. cylindrica*. Land managers have also reported variable *I. cylindrica* control, especially with glyphosate. To determine whether herbicide resistance was a possible explanation, we examined glyphosate response of *I. cylindrica* from 12 Florida populations. We also tested aminocyclopyrachlor with and without glyphosate and flumioxazin with glyphosate. Results indicated that herbicide performance was similar across *I. cylindrica* populations: glyphosate and aminocyclopyrachlor reduced *I. cylindrica* biomass by 78% and 76%, respectively, and the combined tank mix reduced cogongrass biomass by 91%. Flumioxazin tank mixed with glyphosate did not improve control compared with glyphosate alone. There were no differences in pretreatment *I. cylindrica* shoot height, with the exception of two panhandle populations that were shorter. Subsequent harvests indicated few differences in shoot and root plus rhizome weights among untreated controls for almost all populations, with the exception of one of the initially shorter panhandle populations. Our findings indicate that variability in glyphosate efficacy, as suggested by managers, is unlikely due to any conferred resistance. Other abiotic factors such as drought and shade and applicator factors such as carrier water quality should be examined to better understand this issue. Additional studies examining non-target impacts of aminocyclopyrachlor should be conducted to determine its potential fit into *I. cylindrica* management.

## Introduction

Cogongrass [*Imperata cylindrica* (L.) Beauv.] is a C<sub>4</sub> grass from Asia that was accidentally introduced to Mobile, AL, from Japan around 1912 (Dickens 1974). Intentional introductions as a potential forage from material collected in the Philippines occurred in McNeil, MS, in the 1920s and later in Florida in the 1930s and 1940s (Hubbard et al. 1944). Subsequent anthropogenic movements and natural wind-based dispersal have resulted in a widespread distribution across nine southern states. However, the majority of infestations occur in Mississippi, Alabama, and Florida. Impacts in forestry and natural areas have been well documented and continue to be a significant and growing threat (Brewer 2008; Daneshgar et al. 2008; Estrada and Flory 2015).

Management of *I. cylindrica* in forestry and natural areas has been largely herbicide based, as no other effective control strategies have been developed for nonarable lands. Only two herbicides are widely used for *I. cylindrica* management, glyphosate and imazapyr (MacDonald 2004; Minogue et al. 2012). Both can be very effective with repeated applications (Aulakh et al. 2014). However, both may cause significant damage to desirable vegetation (Harmony et al. 2007) from foliar treatments. Imazapyr also exhibits extensive soil residual activity (Czarnota and Derr 2007; Westerman et al. 1993), and many land managers cannot use imazapyr due to non-target injury concerns for many woody and herbaceous plant species (Klaus and Klaus 2011; Kochenderfer et al. 2001; Lewis and McCarthy 2008; Masters et al. 1994).

This leaves a single herbicide, glyphosate, which must be used in an aggressive, repeated manner for effective control. Weed resistance to glyphosate was rare before its widespread use in genetically modified crops (Dyer 1994), but at least 37 species are now glyphosate resistant

© Weed Science Society of America, 2018. This is an Open Access article, distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives licence (<http://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is unaltered and is properly cited. The written permission of Cambridge University Press must be obtained for commercial re-use or in order to create a derivative work.



### Management Implications

Management of cogongrass (*Imperata cylindrica* L. P. Beauv.) is especially challenging, because few effective tools are available for its control. Herbicide treatment remains the recommended approach. However, only two chemicals have proven reliable for *I. cylindrica* management: glyphosate and imazapyr, with the latter often excluded due to non-target impacts. Given the concerns for the development of glyphosate resistance due to repeated application, this research examined glyphosate performance on *I. cylindrica* across 12 total populations from Florida. Four populations were sampled from the western panhandle, from Pensacola to Choctaw Beach, and eight populations were sampled from the peninsular region, from Hilliard to Coral Gables. We also evaluated two other herbicides for potential use in *I. cylindrica* control: aminocyclopyrachlor, both alone and in a tank mix with glyphosate, and flumioxazin in a tank mix with glyphosate. We found limited significant differences in the growth of *I. cylindrica* from different source populations, and the source of the population did not affect herbicide efficacy for either root plus rhizome or total biomass. Aminocyclopyrachlor reduced *I. cylindrica* biomass similar to glyphosate, and the tank mix with both was significantly better than either alone, which warrants additional field testing based on its promising activity in this study. The tank mix of flumioxazin with glyphosate did not significantly improve control compared with glyphosate alone. These results do not suggest that some *I. cylindrica* populations are inherently more difficult to control than others in Florida. Future research should assess the potential influence of environmental variables such as light intensity and moisture stress on glyphosate performance on *I. cylindrica*.

(Heap 2017). Although there are no published studies that have found evidence of *I. cylindrica* tolerance or resistance to glyphosate, sole reliance on glyphosate could lead to resistance selection in *I. cylindrica* with potentially catastrophic consequences. This is especially true for land managers who will not tank mix or rotate to imazapyr due to non-target concerns. Coincidentally, land managers have reported variation in glyphosate performance on *I. cylindrica*. The reasons for this are unclear and warrant further investigation. However, multiple studies have established a high degree of genetic diversity (Burrell et al. 2015; Lucardi et al. 2014) among *I. cylindrica* populations across the southeastern United States.

Aminocyclopyrachlor, a relatively new herbicide labeled for use in non-crop settings, controls a wide range of weeds and invasive plants (Enloe et al. 2015; Minogue et al. 2011). It primarily has activity on eudicots, but some grasses have demonstrated sensitivity to it (Flessner et al. 2011c; Wirt and Lym 2016). Aminocyclopyrachlor functions as an auxin mimic similar to pyridine carboxylic acids such as aminopyralid. Previous research on *I. cylindrica* control with aminocyclopyrachlor has yielded mixed results. Greis (2012) and Enloe et al. (2012) in separate studies in Florida and Alabama, respectively, found aminocyclopyrachlor to show strong suppression of approximately 80% at 6 mo after treatment (MAT), with little to no suppression after 12 MAT. However, Greis (2012) found no reduction in rhizome biomass at 9 or 16 MAT, while Enloe et al. (2012) found a 28% reduction in rhizome biomass at 12 MAT, which was significantly different from the untreated controls.

This may suggest underlying differences in long-term *I. cylindrica* control in south Alabama, near the original Japanese material introduction, versus central peninsular Florida, near the original Philippine material introduction site.

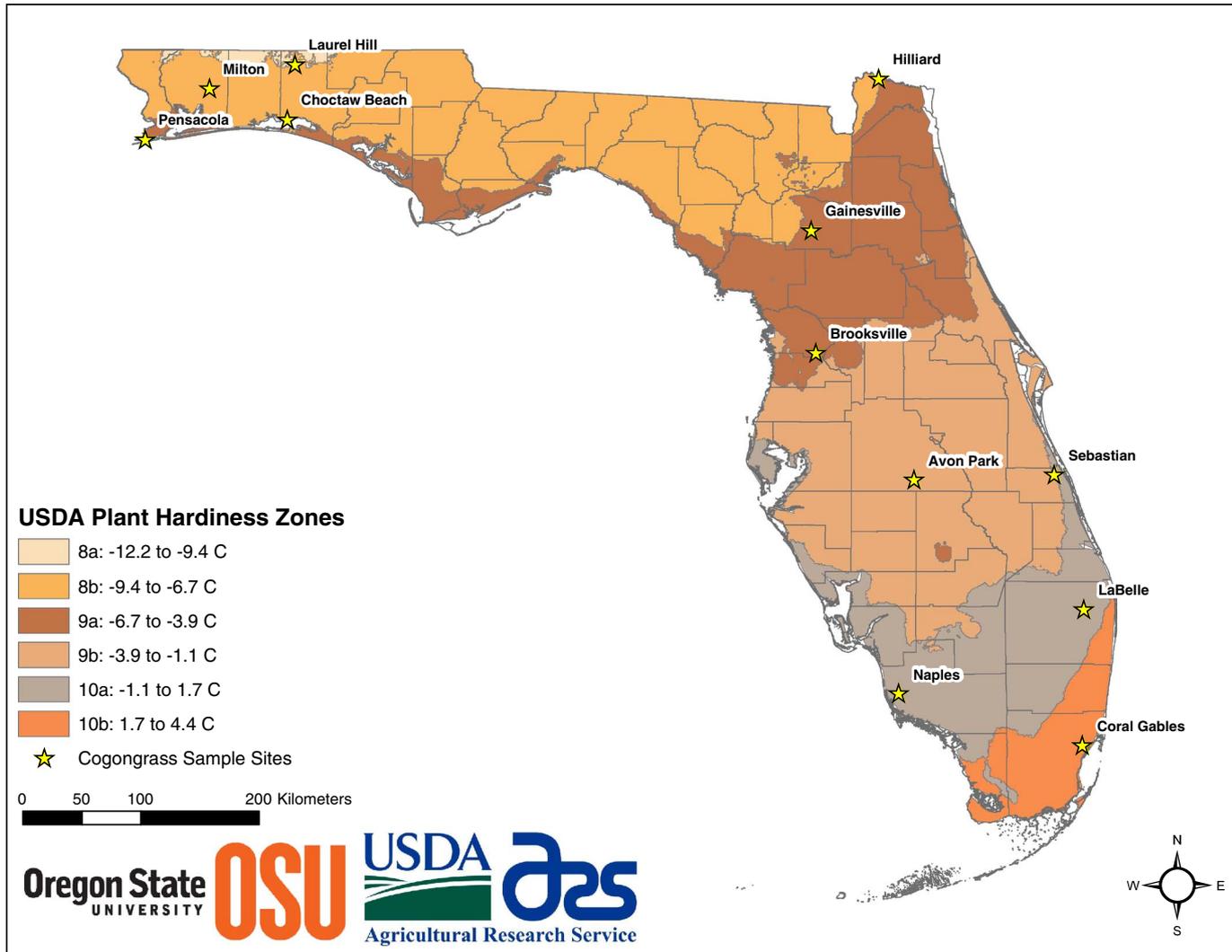
Flumioxazin is a herbicide used in many crop and non-crop settings for control of a wide range of broadleaf and grass weeds. It inhibits protoporphyrinogen oxidase, resulting in rapid membrane disruption and subsequent death in susceptible species (Duke et al. 1991). Flumioxazin has been widely used as a herbicide tank-mix partner in many agronomic studies addressing glyphosate resistance (Beckie 2011), as it provides effective control of many weed species. It is also widely used in aquatic systems, where it controls free-floating species such as water lettuce (*Pistia stratiotes* L.) (Mudge and Haller 2012; Mudge and Netherland 2016). Tank mixes of glyphosate and flumioxazin are also widely used in nursery production and landscape maintenance (Wehtje et al. 2010) due to a lack of impact on many desirable ornamental species. This selectivity might make it a potentially useful tank-mix partner with glyphosate for *I. cylindrica* control.

Given the need for additional tools for *I. cylindrica* control with current concerns for glyphosate resistance, our objective was to evaluate glyphosate and aminocyclopyrachlor as stand-alone treatments and potential tank-mix combinations across a broad range of *I. cylindrica* populations collected throughout Florida. We also evaluated a tank mix of glyphosate and flumioxazin to assess its utility compared with glyphosate alone. We hypothesized that differences in *I. cylindrica* susceptibility to glyphosate from populations sampled throughout Florida would help explain variation in glyphosate performance reported by land managers.

### Materials and Methods

Greenhouse studies were conducted during the spring and summer of 2016 at the University of Florida Center for Aquatic and Invasive Plants in Gainesville, FL. *Imperata cylindrica* rhizomes were collected from 12 populations originating from both the peninsular and western panhandle regions of Florida (Figure 1). Collections encompassed USDA plant hardiness zones 8a in the far northern panhandle near Laurel Hill, FL, to 10b in the southern peninsular region near Coral Gables, FL. Additional details of the collection are provided in Hiatt (2016). Individual rhizomes from each population were planted in 100-L stock pots filled with construction sand. These were fertilized and watered as needed until the new plants were well established and produced sufficient rhizome biomass for experimentation. The process of rhizome propagation for *I. cylindrica* is rapid, and sufficient material was readily generated from the previous growing season. In mid-April 2016, rhizome segments 10 cm in length were harvested from the stock pots. For each population, two rhizome segments were planted into each of 25 (3.8-L) pots for study. Pots were filled with a 1:1 mix of construction sand and potting media (Professional Top Soil, Margo Garden Products, Folkston, GA) mixed with a complete (15-9-12) slow-release fertilizer (Osmocote Plus, Scotts, Maryville, OH). Rhizome segments were planted in each pot at a depth of 2.5 cm. These were watered daily and grown for approximately 75 d until plants were well established. Established plants were those that had produced several new shoots per pot, and new rhizomes were present throughout the pot.

Treatments included glyphosate applied at 0.28 kg ae ha<sup>-1</sup> (Rodeo® herbicide, Dow AgroSciences, Indianapolis, IN), aminocyclopyrachlor applied at 0.28 kg ae ha<sup>-1</sup> (Method® herbicide, Bayer Crop Science, Research Triangle Park, NC), glyphosate + aminocyclopyrachlor



**Figure 1.** Collection locations for the 12 *Imperata cylindrica* populations used in the study. USDA plant hardiness zone map layer accessed with permission at: <http://planthardiness.ars.usda.gov/PHZMWeb> (accessed January 17, 2018).

applied at 0.28 + 0.28 kg ae ha<sup>-1</sup>, glyphosate + flumioxazin (Clipper® herbicide, Valent USA, Walnut Creek, CA ) applied at 0.28 kg ae ha<sup>-1</sup> + 0.14 kg ai ha<sup>-1</sup>, and an untreated control. A nonionic surfactant (Induce, Helena Chemical, Collierville, TN) was added to each herbicide treatment at 0.5% v/v. Treatments were applied with a CO<sub>2</sub>-pressurized backpack boom sprayer equipped with four 8002 flat-fan nozzles at a pressure of 276 kPa and an application volume of 178 L ha<sup>-1</sup>.

Baseline maximum shoot height was measured for each pot (N = 300) just before herbicide treatment. At 30 d after treatment (DAT), shoots were clipped to the soil surface and all live shoots were separated, oven-dried at 65 C for 72 h, and weighed. Plants were then allowed to grow for an additional 30 d. At 60 DAT, all shoot regrowth was harvested as previously described. Additionally, all belowground biomass, which included roots and rhizomes, was harvested, oven-dried, and weighed. The entire experiment was repeated in mid-July 2 wk later in the summer of 2016 on a second set of planted pots and was completed by mid-September. Ambient greenhouse conditions were similar over both experimental runs. The mean daily maximum, mean, and mean minimum temperatures were 32.5, 26.3, and 20.1 C, respectively, for run 1, and 32.4, 26.3, and 20.2 C, respectively, for run 2 (Southern Regional Climate Center accessed January 23, 2018).

**Statistical Analysis**

For each experimental run, five replicate pots for each herbicide by *I. cylindrica* population combination were arranged in a completely randomized design. *Imperata cylindrica* growth by population, as measured by pretreatment shoot height, 30 DAT shoot biomass, and 60 DAT shoot regrowth biomass, was compared using ANOVA for the untreated controls. Additionally, belowground biomass and total biomass (aboveground + belowground biomass) at 60 DAT were compared using ANOVA for all treatments. Herbicide efficacy was compared in terms of a percent reduction in the 60 DAT belowground biomass and total biomass (shoot + belowground) compared with the untreated control. Shoot biomass alone at 60 DAT was not analyzed, because more than 90% of observations were zero following herbicide treatments. This also precluded useful estimation of root to shoot ratios. The analysis of treatment efficacy was performed using the 60 DAT belowground or total biomass averaged for each experimental run, herbicide treatment, and population combination.

Average 60 DAT plant biomass per pot is not a typical continuous variable, because it includes growth, necrosis, and mortality. Other characteristics of this experimental data are that the mass of the untreated control has no defined upper bound, a treatment mean could be zero, treatments have a multiplicative effect with respect to percent reduction, and variance was found to be proportional to the mean. These characteristics lend themselves to analysis methodology applied to count data. Although plant mass is not strictly a count, the analysis of efficacy was modeled using Poisson distributional assumptions with a log-link function as outlined by McCullagh and Nelder (1989) for the biological assay of data from Fisher (1949). The analysis was performed as a generalized linear mixed model using SAS PROC GLIMMIX (Littell et al. 2006). This approach allowed treatments to be compared in terms of percent reduction relative to the untreated control and provided tests of antagonism (see Appendix 1 in the Supplementary Material). The test of antagonism is significant if the difference of tank-mix percent control minus “expected” percent control differs significantly from zero. Expected tank-mix percent

control was calculated as 100 × (1 - (1 - proportion control from herbicide 1 used alone) × (1 - proportion control from herbicide 2 used alone)). Proportion or percent control are not the dependent variables in the analysis or tests of hypotheses, but these results are presented in terms of percent control calculated from biomass treatment means. The rationale for this approach in this analysis is that the difference between a treatment and the untreated check in the linear predictor part of this generalized linear model (that uses a log-link function) equates to treatment biomass divided by the untreated check biomass after applying the inverse link function.

Treatment effects were partitioned using contrast statements to construct a two-way ANOVA for glyphosate and aminocyclopyrachlor herbicides and to test whether glyphosate + flumioxazin and glyphosate + aminocyclopyrachlor tank mixes differed. The test of the aminocyclopyrachlor main effect is that the average of aminocyclopyrachlor and aminocyclopyrachlor + glyphosate treatments differ from the average of the untreated check and glyphosate-alone treatments. The glyphosate main effect is that the average of glyphosate and glyphosate + aminocyclopyrachlor treatments differ from the average of the untreated check and aminocyclopyrachlor treatments. Main effect means were compared using Tukey’s adjustment for multiplicity at P = 0.05 when applicable.

**Results and Discussion**

Pretreatment *I. cylindrica* shoot heights were not different among 9 of the 12 populations and ranged from 104.4 to 123.4 cm (Table 1). Populations from Milton and Pensacola were significantly shorter at 47 and 67.8 cm, respectively. *Imperata cylindrica* from Laurel Hill averaged 89 cm in height and was not different from any other population.

**Table 1.** Comparison of *Imperata cylindrica* panhandle and peninsular Florida population shoot parameters for the untreated controls with means considered normally distributed.

Population	Shoot height <sup>a</sup>	Shoot weight <sup>a</sup>	
	cm	----- g -----	
	0 DAT	30 DAT	60 DAT
Avon Park	123.4 a	16.7 ab	5.4 abc
Brooksville	109.5 a	23.1 a	8.3 a
Choctaw Beach	118.1 a	22.3 a	7.3 ab
Coral Gables	113.6 a	19.4 ab	6.7 abc
Gainesville	114.9 a	17.9 ab	6.3 abc
Hillard	114.5 a	19.3 ab	6.3 abc
LaBelle	104.4 a	15.5 abc	4.0 bc
Laurel Hill	89.0 ab	12.7 abc	5.2 abc
Milton	47.0 c	5.4 c	2.7 c
Naples	104.5 a	14.8 abc	6.1 abc
Pensacola	67.8 bc	9.3 bc	3.7 bc
Sebastian	109.2 a	14.9 abc	4.7 abc
Standard error <sup>b</sup>	10.0	1.9	0.7

<sup>a</sup>Population means within columns followed by the same letter are not significantly different using Tukey’s mean comparison adjustment (P = 0.05).

<sup>b</sup>Standard error of population means from analysis that considers treatment run a random effect.

**Table 2.** Comparison of *Imperata cylindrica* panhandle and peninsular Florida population biomass parameters (standard errors in parentheses<sup>a</sup>) for the untreated controls at the final harvest (60 d after treatment).

Population	Root + rhizome weight <sup>b</sup>	Total weight <sup>b</sup>
	----- g -----	
Avon Park	19.2 (3.5) a	24.6 (3.9) ab
Brooksville	19.2 (3.5) a	27.5 (4.2) ab
Choctaw Beach	23.2 (4.0) a	30.5 (4.5) a
Coral Gables	21.3 (3.8) a	28.1 (4.3) a
Gainesville	19.9 (3.6) a	26.2 (4.1) ab
Hillard	18.7 (3.5) a	25.1 (4.0) ab
Laurel Hill	13.0 (2.8) a	18.2 (3.3) ab
LaBelle	15.2 (3.1) a	19.2 (3.4) ab
Milton	9.7 (2.4) a	12.2 (2.6) b
Naples	16.0 (3.2) a	22.0 (3.7) ab
Pensacola	16.5 (3.2) a	20.2 (3.5) ab
Sebastian	13.2 (2.8) a	17.9 (3.3) ab

<sup>a</sup>Approximate standard error of population means from the generalized linear mixed-model analysis that considers treatment run a random effect.

<sup>b</sup>Population means within columns followed by the same letter are not significantly different using Tukey's mean comparison adjustment ( $P=0.05$ ).

Across all populations, there were also limited differences in *I. cylindrica* shoot biomass for the untreated controls at 30 DAT and for regrowth at 60 DAT (Table 1). There were no significant differences in 10 of the populations for shoot weight at 30 DAT, and these ranged from 12.7 to 23.1 g. Only Milton (5.4 g) produced significantly lower biomass than Brooksville and Choctaw Beach at 30 DAT. Subsequent shoot regrowth at 60 DAT also yielded few differences among populations, as 10 out of 12 populations were not different (Table 1). However, Brooksville had significantly greater regrowth than either Milton or Pensacola. Root plus rhizome weights at 60 DAT for the untreated controls did not differ among any populations and ranged from 9.7 to 23.3 g

(Table 2). Total weight (root + rhizome + shoot regrowth) at 60 DAT did not differ for 11 out of 12 populations and ranged from 17.9 to 30.5 g. Only Milton (12.2 g) was significantly lower than Choctaw Beach or Coral Gables. These pretreatment height data and biomass data from 30 and 60 DAT for the untreated controls indicate similar performance across most populations in our study. Only the Milton population from the panhandle resulted in significantly lower heights and weights than any other population. Explicit reasons for this are unclear but may have been related to possible genetic differences, as Burrell et al. (2015) found a high degree of genetic variation among *I. cylindrica* populations sampled within the panhandle region of Florida.

There were no significant *I. cylindrica* population or population by herbicide treatment interactions ( $P>0.39$  for both root and total biomass) for all herbicide treatments tested at 60 DAT. This was similar to a lack of differences in shoot height and biomass between source populations for the untreated controls. The lack of *I. cylindrica* population either as a significant main effect or in the interaction with herbicide treatment indicates that the herbicide treatments were not influenced by inherent variation among populations tested. This suggests that other factors are contributing to variable *I. cylindrica* control reported by many land managers in Florida.

Across all *I. cylindrica* populations, the herbicide treatment main effect was highly significant for root plus rhizome biomass and total biomass at 60 DAT ( $P<0.001$ ). Glyphosate and aminocyclopyrachlor reduced belowground biomass by 78% and 76%, respectively, and were not significantly different (Table 3). Additionally, the tank mix of these two herbicides reduced belowground biomass by 91% and was significantly greater than either herbicide alone. This increase of 13% to 15% in control would appear promising for improved, local *I. cylindrica* management and increased chemical options for land managers. However, when tests of antagonism were applied to the data, actual control was significantly lower than projected for both belowground biomass and total biomass. This is statistically noteworthy but may be biologically questionable, as the differences were less than 5 percentage points between actual and projected. This pattern was also consistent for total biomass. The tank mix increased percent control by 10% to 11% compared with either herbicide applied alone but was significantly less than projected control (Table 3). Finally, the tank mix of glyphosate + flumioxazin did not improve *I. cylindrica* control compared with

**Table 3.** Comparison of 60 DAT root and total biomass (standard error of means in parentheses) and percent control.

Herbicide	Rate (kg ha <sup>-1</sup> )	Root + rhizome biomass		Total biomass	
		g	% Control <sup>a</sup>	g	% Control <sup>a</sup>
Untreated check	—	16.6 (1.7)	—	22.0 (1.9)	—
Glyphosate	0.28	3.7 (0.5)	78 bc	3.7 (0.5)	83 bc
Aminocyclopyrachlor	0.28	4.0 (0.6)	76 c	4.0 (0.5)	82 c
Glyphosate + aminocyclopyrachlor	0.28 + 0.28	1.6 (0.3)	91 a	1.6 (0.3)	93 a
Glyphosate + flumioxazin	0.28 + 0.14	2.3 (0.4)	86 ab	2.4 (0.4)	89 ab
<i>Test of antagonism</i>					
Glyphosate + aminocyclopyrachlor	0.28 + 0.28		95 <sup>b</sup>		97 <sup>c</sup>

<sup>a</sup>Percent control followed by the same letter does not differ at  $P=0.05$  using Tukey's adjustment for multiplicity.

<sup>b</sup>Expected control was significantly greater than actual control ( $P=0.022$ ).

<sup>c</sup>Expected control was significantly greater than actual control ( $P=0.001$ ).

glyphosate alone for belowground or total biomass. No test of antagonism was performed for the glyphosate + flumioxazin tank mix, since flumioxazin was not included as a stand-alone treatment.

These data indicate a significant impact of aminocyclopyrachlor on *I. cylindrica* biomass, and our data are congruent with other published studies. In field studies conducted in Alabama, Enloe et al. (2012) found aminocyclopyrachlor applied at the same rate in this study ( $0.28 \text{ kg ha}^{-1}$ ) reduced rhizome biomass by 28% at 12 MAT. This rate also reduced vegetative *I. cylindrica* cover by 84% and 54% at 3 and 5 MAT, respectively, but resulted in cover higher than the untreated control at 12 MAT. Additional field dose-response studies reported in Enloe et al. (2012) indicated that aminocyclopyrachlor applied at  $0.56 \text{ kg ha}^{-1}$  resulted in approximately 80% control at 7 to 8 MAT. Similarly, we observed a reduction in *I. cylindrica* belowground and total biomass of near 80% at 60 DAT across our populations.

Although aminocyclopyrachlor primarily controls many broadleaf weeds, certain grasses have also exhibited susceptibility to aminocyclopyrachlor. Parker et al. (2015) found *I. cylindrica* was more susceptible to aminocyclopyrachlor than tall fescue [*Lolium arundinaceum* (Schreb.) Darbysh.], bahiagrass (*Paspalum notatum* Fluegg), and bermudagrass [*Cynodon dactylon* (L.) Pers.]. Additionally, Flessner et al. (2011b) found differential susceptibility among six Japanese lawngress (*Zoysia japonica* Steud.) cultivars to aminocyclopyrachlor. The physiological basis for sensitivity in certain grasses is not completely understood. However, in St. Augustinegrass [*Stenotaphrum secundatum* (Walter) Kuntz], Flessner et al. (2011a) found injury consistent with other synthetic auxin-type herbicides that included growth stimulation and vascular inhibition.

Our data demonstrated consistent susceptibility to aminocyclopyrachlor in both panhandle and peninsular *I. cylindrica* populations from across a wide geographic range in Florida. One major consideration with aminocyclopyrachlor that needs to be addressed is potential non-target injury in natural areas. Aminocyclopyrachlor has resulted in injury to non-target species through soil residual activity (Kniss and Lyon 2011) and release from treated plants (Lewis et al. 2013). Although not classified as purely nonselective, a better understanding of nontarget impacts in southeastern ecosystems where *I. cylindrica* is prominent is critical to developing potential use patterns.

Regarding the glyphosate + flumioxazin tank mix, we found no evidence to suggest flumioxazin improved *I. cylindrica* control with glyphosate. The tank mix resulted in an 89% reduction in total biomass, while glyphosate alone resulted in an 83% reduction, and these were not significantly different. Byrd (2007) tested flumioxazin at 0.07, 0.14, and  $0.28 \text{ kg ha}^{-1}$  with ammonium sulfate at  $0.51 \text{ kg ha}^{-1}$  and glyphosate at  $1.68 \text{ kg ha}^{-1}$ . He reported no difference in *I. cylindrica* control when glyphosate was applied with or without any rate of flumioxazin. Glomski (2013) examined flumioxazin tank mixes with glyphosate for control of Japanese knotweed (*Reynoutria japonica* Houtt.) at 0.05, 0.11, and  $0.21 \text{ kg ha}^{-1}$  with and without  $2.0 \text{ kg ha}^{-1}$  glyphosate. In that study, they reported no difference in treatment performance when flumioxazin was applied alone or with glyphosate. Our study found a comparable outcome, as flumioxazin was not an effective tank additive.

In conclusion, our findings suggest that variability in efficacy, as suggested by managers, is unlikely due to conferred resistance. Other abiotic factors such as drought, shade, and seasonality of treatment may be important. Additionally, water quality, which is well known to impact glyphosate performance, may be important (Nalewaja and Matysiak 1991). Variation in herbicide application

methods for broadcast- and spot-treatment approaches should also be examined, as they likely vary considerably in dose among applicators, especially for spot treatments. While no indication of glyphosate tolerance or resistance was observed, the ecological implications of detected growth differences among some *I. cylindrica* populations, albeit minor, also indicates a need for additional studies to better understand the invasion ecology of *I. cylindrica* in the southeastern United States. Finally, aminocyclopyrachlor, which demonstrated considerable activity on *I. cylindrica*, should be further examined, especially from a non-target perspective. Identifying the susceptible species range to determine potential collateral impacts of this active ingredient in a resistance management rotation would be very useful.

**Supplementary material.** To view supplementary material for this article, please visit <https://doi.org/10.1017/inp.2018.12>

**Acknowledgments.** The authors would like to thank Drew Hiatt for collecting and sharing all *I. cylindrica* populations used in this study. This research was supported by the University of Florida Center for Aquatic and Invasive Plants and by the USDA Forest Service Southern Research Station (14CA1130129051).

## References

- Aulakh JS, Enloe SF, Loewenstein NJ, Price AJ, Wehtje G, Miller JH (2014) Pushing towards cogongrass patch eradication: the influence of herbicide treatment and application timing on cogongrass rhizome elimination. *Invasive Plant Sci Manag* 7:398–407
- Beckie HJ (2011) Herbicide-resistant weed management: focus on glyphosate. *Pest Manag Sci* 67:1037–1048
- Brewer S (2008) Declines in plant species richness and endemic plant species in longleaf pine savannas invaded by *Imperata cylindrica*. *Biol Invasions* 10:1257–1264
- Burrell AM, Pepper AE, Hodnett G, Goolsby JA, Overholt WA, Racelis AE, Diaz R, Klein PE (2015) Exploring origins, invasion history and genetic diversity of *Imperata cylindrica* (L.) P. Beauv. (Cogongrass) in the United States using genotyping by sequencing. *Mol Ecol* 24:2177–2193
- Byrd JD Jr (2007) What works on cogongrass and what does not: a summary of nearly 10 years of cogongrass research in Mississippi. Pages 28–31 in Loewenstein NJ & Miller JH, eds., *Proceedings of the Regional Cogongrass Conference: A Cogongrass Management Guide*, Mobile, AL. Auburn, AL: Alabama Cooperative Extension System
- Czarnota M, Derr J (2007) Controlling Bamboo (*Phyllostachys* spp.) with herbicides. *Weed Technol* 21:80–83
- Daneshgar P, Jose S, Collins A, Ramsey C (2008) Cogongrass (*Imperata cylindrica*), an alien invasive grass, reduces survival and productivity of an establishing pine forest. *For Sci* 54:579–587
- Dickens R (1974) Cogongrass in Alabama after sixty years. *Weed Sci* 22:177–179
- Duke SO, Lydon J, Becerril JM, Sherman TD, Lehnen LP Jr, Matsumoto H (1991) Protoporphyrinogen oxidase-inhibiting herbicides. *Weed Sci* 39:465–473
- Dyer WE (1994) Resistance to glyphosate. pp. 229–241 in Powles S & Holtum J, eds., *Herbicide Resistance in Plants: Biology and Biochemistry*. New York: CRC Lewis
- Enloe SF, Belcher J, Loewenstein NJ, Aulakh JS, van Santen E (2012) Cogongrass control with aminocyclopyrachlor in pastures. forage and grazinglands. Plant Management Network. <https://dl.sciencesocieties.org/publications/fg/pdfs/10/1/2012-0828-02-RS>. Accessed: August 15, 2017
- Enloe SF, Loewenstein NJ, Streett DA, Lauer DK (2015) Herbicide treatment and application method influence root sprouting in Chinese tallowtree (*Triadica sebifera*). *Invasive Plant Sci Manag* 8:160–168
- Estrada JA, Flory SL (2015) Cogongrass (*Imperata cylindrica*) invasions in the US: mechanisms, impacts, and threats to biodiversity. *Global Ecol Conserv* 3:1–10
- Fisher RA (1949) A biological assay of tuberculins. *Biometrics* 5:300–316
- Flessner M, Dute R, McElroy J (2011a) Anatomical response of St. Augustinegrass to aminocyclopyrachlor treatment. *Weed Sci* 59:263–269

- Flessner ML, McCurdy JD, McElroy JS (2011b) Tolerance of six zoysiagrass cultivars to aminocyclopyrachlor. *Weed Technol* 25:574–579
- Flessner ML, McElroy JS, Wehtje GR (2011c) Quantification of warm-season turfgrass injury from triclopyr and aminocyclopyrachlor. *Weed Technol* 53:367–373
- Glomski LM (2013) Herbicide Combinations for Controlling Japanese Knotweed. Vicksburg, MS: U.S. Army Engineer Research and Development Center APCRP ERDC/TN APCRP-CC-19, Technical Notes Collection. <https://erdc-library.ercd.dren.mil/xmlui/handle/11681/3842>. Accessed: April 16, 2018
- Greis AL (2012) Characterization and Evaluation of Aminocyclopyrachlor on Native and Invasive Species of Florida. Master's thesis. University of Florida. 103 p
- Harmony K, Stahlman P, Hickman K (2007) Suppression of Caucasian Old World bluestem with split application of herbicides. *Weed Technol* 21:573–577
- Heap I (2017) The International Survey of Herbicide Resistant Weeds. [www.weedscience.org](http://www.weedscience.org). Accessed: August 1, 2017
- Hiatt D (2016) Invasive Plant Populations and Co-occurring Native Species Vary in Phenotypic Plasticity. Master's thesis. University of Florida. 46 p
- Hubbard CE, Gray AP, Brown D, Whyte RO (1944) *Imperata cylindrica*: Taxonomy, Distribution, Economic Significance, and Control. Oxford, U.K.: Imperial Agricultural Bureaux Joint Publication 7. 63 p
- Klaus NA, Klaus JM (2011) Evaluating tolerance of herbicide and transplantation by cane (a native bamboo) for canebrake restoration. *Restor Ecol* 19:344–350
- Kniss A, Lyon D (2011) Winter wheat response to preplant applications of aminocyclopyrachlor. *Weed Technol* 25:51–57
- Kochenderfer JD, Zedaker SM, Johnson JE, Smith DW, Miller GW (2001) Herbicide hardwood crop tree release in central West Virginia. *North J Appl For* 18:46–54
- Lewis DF, Richardson RJ, Yelverton FH, Wentworth TR (2013) Bioavailability of aminocyclopyrachlor and triclopyr plus clopyralid from turfgrass clippings in aquatic and riparian plants. *Weed Sci* 61:594–600
- Lewis K, McCarthy B (2008) Nontarget tree mortality after tree-of-heaven (*Ailanthus altissima*) injection with imazapyr. *North J Appl For* 25:66–72
- Littell RC, Milliken GA, Stroup WW, Wolfinger RD, Schabenberger O (2006) SAS® for Mixed Models. 2nd ed. Cary, NC: SAS Institute. 814 p
- Lucardi RD, Wallace LE, Ervin GN (2014) Invasion success in Cogongrass (*Imperata cylindrica*): a population genetic approach exploring genetic diversity and historical introductions. *Invasive Plant Sci Manag* 7:59–75
- MacDonald GE (2004) Cogongrass (*Imperata cylindrica*)—biology, ecology, and management. *Crit Rev Plant Sci* 23:367–380
- Masters R, Stougaard R, Nissen S (1994) Leafy spurge (*Euphorbia esula*) control with fall-applied imazapyr, imazaquin, and imazethapyr. *Weed Technol* 8:58–63
- McCullagh P, Nelder JA (1989) Generalized Linear Models. Chapman & Hall/CRC New York. 511 p
- Minogue P, Enloe S, Osiecka A, Lauer D (2011) Comparison of aminocyclopyrachlor to common herbicides for kudzu (*Pueraria montana*) management. *Invasive Plant Sci Manag* 4:419–426
- Minogue PJ, Miller JH, Lauer DK (2012) Use of glyphosate and imazapyr for cogongrass management in southern pine forests. *South J Appl For* 36:19–25
- Mudge C, Haller WM (2012) Response of target and nontarget floating and emergent aquatic plants to flumioxazin. *J Aquat Plant Manage* 50:111–116
- Mudge C, Netherland MD (2016) Continued Development of Use Patterns for Combining Glyphosate and Protox Inhibiting Herbicides for Control of Invasive Emergent and Floating Aquatic Plants. Final Report. Tallahassee, FL: Florida Fish and Wildlife Conservation Commission Research Grant Program. 11 p
- Nalewaja JD, Matysiak R (1991) Salt antagonism of glyphosate. *Weed Sci* 39:622–628
- Parker E, Wehtje G, McElroy J, Flessner M, Panizzi P (2015) Physiological basis for differential selectivity of four grass species to aminocyclopyrachlor. *Weed Sci* 63:788–798
- Southern Regional Climate Center. CLIMDAT Data Tool. <https://climdata.srcc.lsu.edu>. Accessed January 23, 2018
- Wehtje G, Gilliam C, Marble S (2010) Postemergence weed control with glyphosate plus flumioxazin combinations. *Weed Technol* 24:356–360
- Westerman R, Murray D, Castner E (1993) Hogpotato (*Hoffmanseggia glauca*) control with herbicides and rotational crop response. *Weed Technol* 7: 650–656
- Wirt K, Lym R (2016) Effect of aminocyclopyrachlor on seedling grasses. *Invasive Plant Sci Manag* 9:87–95