

ADVANCED PHENOTYPING USING FT-IR DISTINGUISHES DISEASE RESISTANCE IN *FRAXINUS EXCELSIOR* AGAINST *HYMENOSCYPHUS FRAXINEUS*

Caterina Villari¹, Arnaud Dowkiw², Rasmus Enderle³, Marjan Ghasemkhani⁴, Thomas Kirisits⁵, Erik Kjaer⁶, Diana Marčiulyrienė⁷, Lea McKinney⁶, Berthold Metzler³, Facundo Muñoz², Lene Rostgaard Nielsen⁶, Alfaf Pliūra⁷, Lars-Göran Stener⁸, Vytautas Suchockas⁷, Luis Rodriguez-Saona⁹, Pierluigi Bonello¹⁰, and Michelle Cleary⁴

The invasive ascomycete fungus *Hymenoscyphus fraxineus* has caused large scale population decline of European ash (*Fraxinus excelsior*) throughout much of its distribution range in Europe. Large genotypic variation in susceptibility to the pathogen exists in the natural population, albeit resistance occurs at a low level. Damage traits associated with the pathogen are strongly genetically controlled, which suggests that considerable gain can be achieved through selection and breeding. Large potential exists for utilizing modern approaches or tools that can quantitatively identify phenotypes and help explain the genetic basis of important plant traits, such as disease resistance. One main obstacle to making such advances is the lack of viable tools for rapid resistance phenotyping. Vibrational spectroscopy is one approach that has been used successfully for rapid phenotyping for plant resistance. In this study, we used Fourier-transform Infrared (FT-IR) spectroscopy coupled with a chemometric model to discriminate between resistant and susceptible European ash genotypes. Non-infected leaves and stem samples from known susceptible and resistant *F. excelsior* genotypes were collected from six European countries. Purified phenolic extracts were analyzed on a FT-IR spectrometer and soft independent modeling of class analogy (SIMCA) was used to discriminate between resistant and susceptible trees. The model built with stem samples, and subsequently validated using random blind samples, gives powerful evidence that FT-IR can clearly discriminate between susceptible and resistant genotypes. These results suggest that spectroscopic phenotyping tools may allow for superior genotypes to be quickly identified and employed in restoration efforts.

¹Warnell School of Forestry & Natural Resources, University of Georgia, Athens, GA 30602.

²Institut National de la Recherche Agronomique (INRA) 2163 Avenue de la Pomme de Pin, 45075 Ardon, Orléans, France.

³Forest Research Institute BadenWuerttemberg, Department Forest Protection Wonnhaldestrasse 4, 79100 Freiburg, Germany.

⁴Swedish University of Agricultural Sciences (SLU), Southern Swedish Forest Research Centre Sundsvägen 3, 23053 Alnarp, Sweden.

⁵Institute of Forest Entomology, Forest Pathology and Forest Protection (IFFF), Department of Forest and Soil Sciences, University of Natural Resources and Life Sciences, (BOKU), Peter-Jordan-Straße 82, 1190 Vienna, Austria.

⁶Department of Geosciences and Natural Resource Management, University of Copenhagen Rolighedsvvej 23, 1958 Frb Copenhagen, Denmark.

⁷Lithuanian Research Centre for Agriculture and Forestry, Institute of Forestry Liepu 1, LT53101 Girionys, Kaunas district, Lithuania.

⁸SKOGFORSK - The Forest Research Institute, Ekebo 2250, 26890 Svalöv, Sweden.

⁹Department of Food Science and Technology, Ohio State University, Parker Food Science and Technology Columbus, OH 43210.

¹⁰Department of Plant Pathology, Ohio State University, 201 Kottman Hall Columbus, OH 43210, (Michelle.Cleary@slu.se).