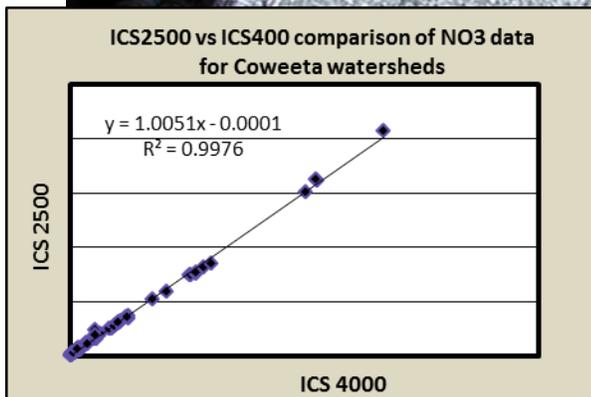
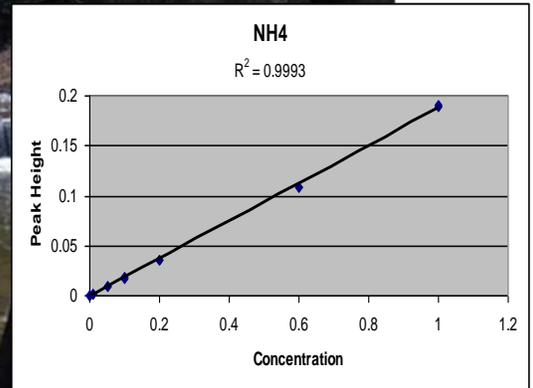
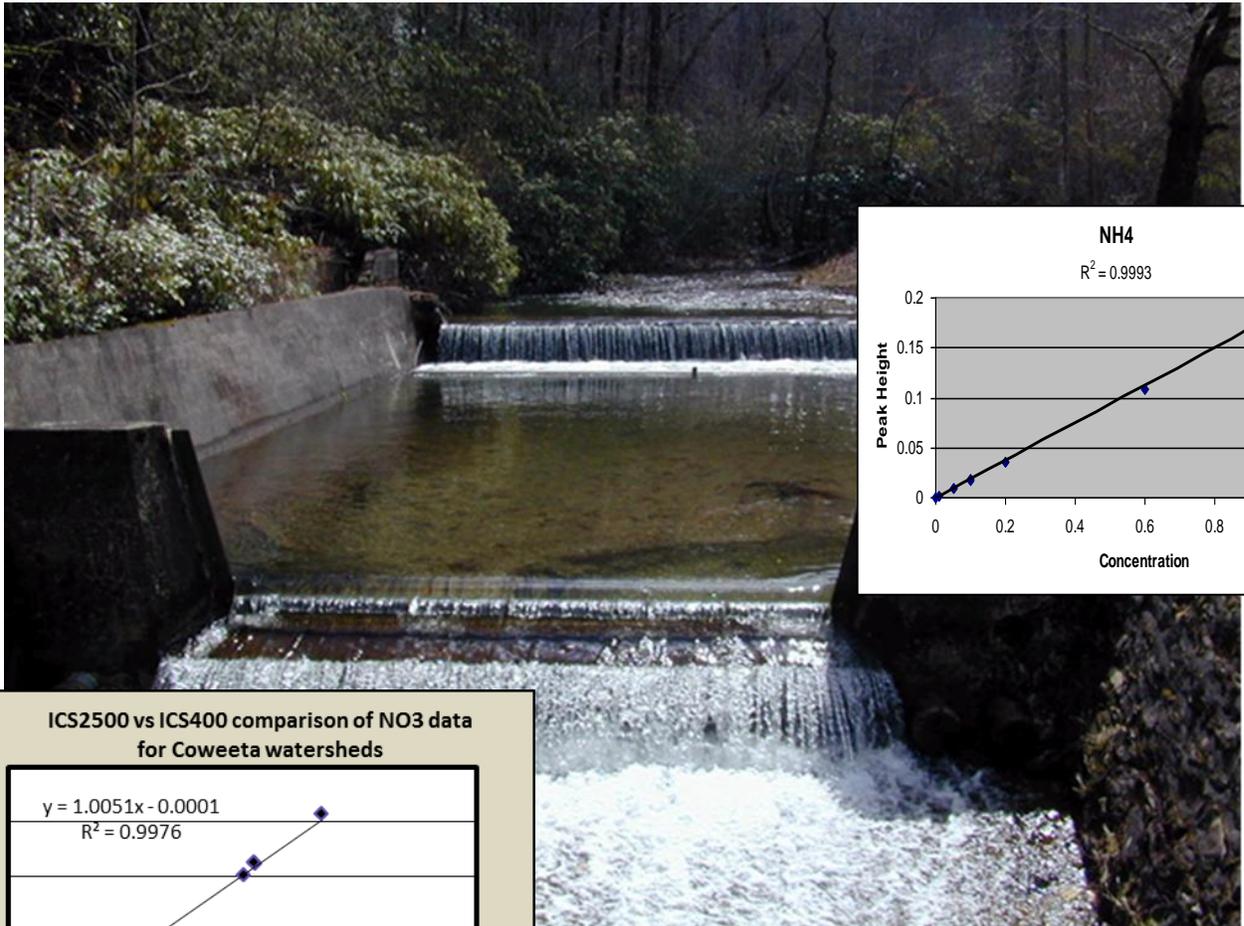


Coweeta Hydrologic Laboratory

QUALITY ASSURANCE PROTOCOL

Revised March 25, 2014



The management of Coweeta Hydrologic Laboratory is fully committed to the maintenance of an effective quality assurance program in order to preserve the integrity of all data collected and reported at Coweeta. Within the quality assurance program are quality control procedures for each analysis.

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Project Leader

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03/24/14

Date

All analyses performed at Coweeta must follow quality assurance guidelines. The following are sample types analyzed at Coweeta.

Water:

Watershed

Rain gauge

Lysimeter

Stream grab

Stream

Sigma stream samples

Overland flow

Wells

Soils

Plant Tissue:

Roots

Leaves

Wood

Tree increment cores

Litterbag

Forest Floor

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Quality Assurance Requirements

I. Field Measurements, Sample Collection, Sampling Frequency and Chain of Custody

A. Field Measurements and Sample Collection

1. A field measurement or collected sample must be representative of the parameter or material that is to be analyzed.
2. The primary investigator assigns the sample identity (ID).
3. Field sampling should include procedures to prevent cross contamination.
4. Replicate samples are collected for heterogeneous material such as soil and forest floor as dictated by the study plan.
5. Before placed out in the field, lysimeters are cleaned with 10% HCl and rinsed with deionized water (DI) until the conductivity is below $5\mu\text{mho/cm}$. The lysimeter is shown to be stable before data are recorded. Repeated sampling and analysis accomplish this and the results are then recorded.

B. Sampling frequency is dictated by the research objectives as defined in the study plan.

C. A Request for Analytical Services (see Appendix A) must be completed by the investigating scientist, authorized by the project leader and presented to the lab manager before samples can be analyzed.

D. Chain of Custody

1. Scientist submits samples to the lab.
2. The Project Leader assigns sample priority.
3. The Lab Manager assures sample flow and disposition.

II. Preparation, Labeling, and Storage of Samples

A. Some samples require the addition of a preservative to increase the shelf life of the sample. See Appendix B for a list of sample matrix type and shelf life. See Appendix C for a list of preservation techniques and maximum holding times for a number of parameters.

B. Field technicians assure proper labeling of the sample, which include sample ID, date of collection, and the name of the principal investigator.

C. Stream, lysimeter and collected water samples are stored in the Walk in Cooler. The cooler is kept at a temperature of $4^{\circ}\text{C} \pm 2^{\circ}\text{C}$. Perishable samples, defined as samples not treated with a preservative, are run within the specified shelf life (see Appendix B). If the sample cannot be analyzed within the specified shelf life the sample is frozen at -18°C .

D. Extracted NH_4Cl , KCl , NH_4Ac and double acid samples, can be unstable. Therefore, the shelf life is determined and documented (see Appendix B). The samples are run before the shelf life is exceeded.

E. The specific shelf life will be determined for all new sample matrices. See Appendix B.

F. Soil samples are stored in the Walk in Cooler.

G. Soil samples to be dried are stored in the insulated attic above the lab.

H. Extracted soil samples are stored in the refrigerator nearest to the lab in which analysis will be performed (i.e. extracted samples for cation analysis are stored in the refrigerator located in the ICP lab).

I. Plant tissue samples are stored in air tight glass vials.

J. Samples to be archived are documented on a spreadsheet and given an UPC code then transferred to the Coweeta sample vault.

III. Laboratory Water Quality

Samples at Coweeta are characterized by very low concentrations of nutrients. The mean concentration for NO₃-N in a control watershed in 1992 was 6µg/L. The need for quality water cannot be overstated.

Deionized Water –A Millipore Milli-Q Integral system is used to give two types of water, Type I (ultra-pure) and Type II which meet or exceed standards set by the American Society of Testing and Materials. The system has a 200 liter storage tank that uses UV to eliminate biological activity in the tank. The Ultra-pure water produced by the Milli-Q is monitored by the DI system and maintains a resistivity of 18.2megaohm cm. Type II has a resistivity of 1-10 megaohm cm. It is monitored through conductivity and pH checks by a lab technician and is documented on the Quality Assurance spreadsheet. The conductivity of type II water should not exceed 1µS/cm.

IV. Labware

A. Glass and Plastic Ware: Sample containers

1. Polyethylene bottles and caps or 50ml polypropylene tubes are used for water sample collection and storage. See Appendix C for required containers, preservation techniques and maximum holding times for a number of parameters.
2. Lab personnel should be familiar with glassware limitations. Graduated cylinders are labeled TC or TD, to contain or to draw. This means liquid left behind in a cylinder labeled TC is part of the volume. The volume from cylinders labeled TD is not dependent on what is left behind. Volumetric's differ in accuracy. A red line volumetric is most accurate and therefore is used for calibration standards or QC standard preparation.
3. Sample containers are discarded if cracks or inside scratches are present. All sample tubes are discarded after one-year use.

B. Washing protocol

1. All labware is washed using Liqui- Nox®. All labware is rinsed 5 times using Type II DI water.
2. Some vials, glassware and sample tubes are rinsed in acid solution. See Procedures for Chemical Analysis, Washing of Glassware and Bottles under Laboratory Protocol for further information.

V. Quality Control Standards, Reagents, Solvents, Acids, and Bases

A. Quality Control and Calibration Standards

1. Certified quality control (QC) standards are purchased from NSI solutions, Environmental Resource Associates (ERA) or a reputable company and are run on all instruments quarterly. The acceptable range is given by the manufacturer.
2. Reference standards are certified standards purchased from reputable commercial vendors such as National Institute of Standards and Technology (NIST).
3. Calibration standards are made up using good laboratory practices from certified stock standards using Type I DI water. QC standards are used to verify calibration curves generated using calibration standards. Calibrants are evaluated quarterly using NSI certified quality control standards or certified standards purchased from a reputable vendor.

B. Chemical Materials

1. The quality of the material must coincide with procedure needs; i.e. ICP work must have spectral grade. For most other applications, analytical grade reagents, solvents, acids and bases are purchased.

2. An inventory spreadsheet is available on the Lab Managers computer. The inventory spreadsheet identifies all chemicals used at Coweeta.
3. All containers are marked with date received and date opened. These are included on the inventory spreadsheet along with date of expiration, vendor, and storage requirements. The inventory spreadsheet includes the method for responsible disposal for expired material. This sheet is checked yearly by the QA officer.

VI. Procedure for Quarterly QC Analysis

- A. When QC samples arrive from the vendor, the QA officer enters the certified values on the quarterly QC spreadsheet, located on the lab manager's computer. Each instrument has a page in the spreadsheet containing calculated values for dilution. The dilutions reflect the value covering the instrument range.
- B. Each analyst makes up a stock solution per the manufacturers instructions. Subsequent dilutions are made from the stock.
- C. All stock solutions and dilutions are made using a red line volumetric and Type I DI water.
- D. If values fall outside the range, the analyst reruns the samples and if still a problem, remakes the QC's.
- E. If the values are still outside the range the QA officer and lab manager are informed. The QA officer assists in determining the problem.

VII. Chemical Analyses

- A. Approved procedures are documented in the Procedures for Chemical Analysis. The Procedures for Chemical Analysis is updated as needed by the Lab Manager.
- B. All analysis performed on an instrument require a standard curve using approved calibrants. The correlation coefficient for the calibration curve must be 99% or better. A Quality Control sample is used to verify the curve and is included in the run as specified by the method. The QC sample must fall within the established range. If the value falls outside the range the data are unacceptable. The problem is isolated and corrected before continuing. Samples are rerun to the point where the problem occurred.
- C. Three reference samples plus three blanks are included in every extraction and digestion. The percent error allowed for a reference sample is specified by the manufacturer. If the value falls outside this range the data are unacceptable. The problem is isolated and corrected before continuing. Samples are rerun to the point where the problem occurred.
- D. The matrix of the calibration standards, reference standard and blanks used in a run match the matrix of the sample; unless it has been determined the analysis is not affected by the matrix.
- E. Dilutions
 1. Automatic pipettes are checked quarterly and logged on the QC spreadsheet by the QA officer.
- F. All dilutions made by hand are done in duplicate and must agree within 10%.
- G. Modifications to a procedure in the Procedures for Chemical Analysis require the approval of the Project Leader.
 1. To modify a procedure, the new procedure and old procedure must be run using the same samples. If the sample is extracted or digested, then the same extracted or digested sample should be used for both procedures. This must be done for all sample matrices used with the procedure.
 2. The results are compared and if R^2 is not 0.95 or greater with a slope of 1, then a correlation factor is developed using regression analysis. All subsequent data is then corrected to a single procedure using the correlation equation.

- H. New procedures should be tested to include correlation with other methods. The Project Leader must approve any new procedure.
- I. After approval of a modified method or acceptance of a new method the lab manager updates the Procedures for Chemical Analysis.
- H. New Sample Studies
 - New sample sets require verification the proper procedure is used for all conditions that affect the matrix of the sample.
 - 1. The QA officer determines if there is a matrix problem using the method of standard additions or another instrument type when available.
 - 2. If a change in procedure of analysis is required, then the following applies:
 - a. The QA officer develops new procedure for the sample set
 - b. The new procedure (including report explaining problem, data comparisons) is sent to the lab liaison and project leader for approval
 - c. The lab manager receives the new procedure for the cookbook
 - 3. The lab manager has the primary investigator sign off on the first sample set analyzed.

VIII. Trouble Shooting Problem with Analysis

When a problem with an analysis occurs (such as a matrix problem) then a procedure to identify and fix the problem must be established. This entails first isolation of questionable data and then development of a new procedure of analysis to deal with the samples. The QA officer determines the analysis to be undertaken. This should include analysis to justify the procedure such as the method of standard additions or samples run on an appropriate instrument such as the IC for comparison with the AutoAnalyzer.

When data/analysis problems are realized the following steps are taken:

1. Notify QA officer and lab manager
2. The lab manager isolates the samples and data in question
3. The QA officer and lab manager develop new procedures for the sample set, either through preparation or analysis
4. The new procedure (including report explaining problem, data comparisons) to the lab liaison and project leader for approval
5. The lab manager receives the new procedure for the cookbook
6. Disposition of questionable data is made by the project leader

IX. Blanks

Analytical blanks or experimental blanks are included in all procedures for extracted and digested samples. A DI blank is included in the weekly Dryfall Collector analysis.

X. Blind samples

Principle investigators are encouraged to include blind checks, duplicates, and blanks with their samples. The QA officer should present blind samples for analysis to each analyst quarterly.

XI. Analysts, training

A qualified Coweeta technician trains new laboratory staff. The Coweeta technician works closely with that person until he/she becomes proficient with an analysis and/or instrument.

XII. Instrumentation

A. Maintenance

1. Each instrument analyst carries out routine maintenance and records this information in a logbook.

2. All logbook maintenance information is transferred to a spreadsheet located on the Lab Managers computer.
 3. Laboratory balances are professionally cleaned and calibrated once a year by a qualified outside technician.
- B. Quarterly quality control checks are made on each instrument using purchased certified quality control standards. The results are recorded on the Quality Control spreadsheet. The information for each calibration curve is included on the Quality Control spreadsheet. The percent error for each instrument is found in Appendix D at the end of this document. The vendor certifying the quality control sample has a certified value and a range associated with that value. This range is the allowable percent error. These values are based on actual historical data collected by the vendor. The range reflects any bias in the method used to establish the limits and closely approximate a 95% confidence interval of the performance that experienced laboratories should achieve using acceptable environmental methods. If results are outside of the allowable error, then the cause is documented and resolved before more data is collected. Results obtained between the current Quarterly check and the previous Quarterly check is scrutinized and inferior data is reported to the QA Officer for disposition.
- C. Instrument background noise is evaluated during each run.
- D. Lower limits of detection, instrument detection limit and method detection limits are defined for each instrument in the Procedures for Chemical Analysis. Methods Detection Limits can be found in Appendix E in this document. These are updated yearly.
- E. New Instrument
- When a new instrument is purchased, the instrument accuracy, precision and stability must be determined. Before the old model is replaced the following is run on both instruments:
1. Run quarterly quality control.
 2. After passing the above, run all sample types on both instruments for comparison.
 3. The data is then reviewed by the lab liaison and project leader to replace the old instrument with the new instrument.

XIII. Data Storage and Reporting

- A. Significant figures are defined to three significant figures except for pH and, soil and litter weights. These are defined to two places.
- B. Below the detection limit values and less than zero designations should not be substituted for the actual value. Actual results obtained including negative numbers should be recorded. See Waite et al. (1980)¹ and Gilbert and Kinnison (1981)² for techniques in averaging data sets containing the above. Each investigator must determine the appropriate method for those data.
- C. Permanent records are maintained for each instrument in a logbook, which include:
 1. Date samples were run, data file ID, and samples ID.
 2. Routine maintenance and major service repairs.
 3. Unusual occurrences during the run.
 4. Logbook information containing maintenance checks, instrument problems and service repairs are entered on the Sample Count Spreadsheet located on the Lab Managers computer.

¹ Waite, D.A., D.H. Denham, J.E. Johnson, D.E. Michels and N. Turnage "Statistical Methods for Environmental Radiation Data Interpretation" J.E. Watson (chairman) in: *Upgrading environmental data* Health Physics Society Committee Report HPSR-1, pp7-1 to 7-19, U.S. EPA Report EPA 520/1-80-012(1980)

² Gilbert, R.O. and R.R. Kinnison "statistical Methods for Estimating the Mean and Variance from Radionuclide Data Sets containing Negative, Unreported or Less-Than values" Health Physics ,40, 377-390(1981)

D. Disposition of data

1. Each analyst examines data before entering on Analytical Lab spreadsheet to ensure calibrants and QC values are good.
2. After entering data on the appropriate work sheet each analyst checks to ensure data is reasonable and agrees with historical data. Some spreadsheets contain conditional formatting for historical values of the sample. Conditional formatting gives the value in red rather than black if the value is outside historical data values. The analyst should then rerun that sample. If all values are red or out of the range from historical values, then a problem could exist and must be evaluated. If it is determined the data is questionable then the procedure outlined in troubleshooting problems with analysis is used.
3. After all analyses have been run, the lab manager checks the data for anomalies and errors. Once checked the data is copied to the appropriate folder. The data in this folder is then ready for use outside the lab.

E. Data files sent to the Lab Manager's computer are backed up every Friday morning on a backup drive. The backup drive is stored in the Coweeta vault.

XIV. Facilities

- A. Heating, cooling ventilation and utilities are maintained to provide a consistent controllable environment.
- B. Electricity, compressed gasses and water are of appropriate quality to meet QC standards.
- C. Laboratory Cleanliness
 1. Soils, litter and forest floor samples should be weighed in the soil lab or back room.
 2. All floor areas should be swept following sampling preparation if necessary.
 3. Balances, lab bench tops and hoods should be kept free of chemicals. All chemical spills should be cleaned immediately.
 4. Because fixtures such as lights, spigots and air taps corrode they should be checked yearly.

Appendix A

WETLAB - REQUEST FOR ANALYTICAL SERVICES

Name of Study _____

Principle Investigator _____

Date(s) Sampling Initiated _____

Type of sample (i.e. Water, Soil, Plant) _____

A. Number of samples per collection _____

Sample ID's _____

B. Requested completion date _____

Sample Shelf Life _____

Project Leader Approval _____

Water	Soil	Plant Tissue
NH ₄ -N	SOIL EXTRACTIONS:	TOTAL CATIONS
NO ₃ -N	1) KCL (NO ₃ -N, NH ₄ -N)	CHN
Br	2) DBL. ACID :	
SO ₄	cations	
Cl	P	ARCHIVE
SIO ₂	3) NH ₄ CL:	
O-PO ₄	cations	
K	CEC	
Na	4) Phosphate extract (SO ₄)	
Ca		
Mg	TOTAL CATIONS	
Al	SOIL BULK DENSITY	
pH	SOIL MOISTURE	
HCO ₃	SOIL pH	
TOTAL N (TKN)	EX. ACIDITY	
TP	CHN	
CONDUCTIVITY	PRESSURE PLATE	
DOC/TN		
TSS	ARCHIVE	

Check beside each desired analysis *:

*For additional information on analytical methods refer to:

“Procedures for Chemical Analysis Coweeta Hydrologic Lab”

“Standard Methods for the Examination of Water”

Consult with Cindi Brown – Lab Manager

Appendix B –Shelf Life

Shelf Life¹

**	SO4	o-PO4	TP*	NO3	Cl	Br	NH4	SiO3	K	Na	Ca	Mg	DOC	TN	pH
H2O	5 days	2 days	2 days	2 days	5 days	28 days	2 days	28 days	2 weeks	2 weeks	2 weeks	2 weeks	5 days	5 days	Immediately
NH4Cl	na	na	na	na	na	na	na	na	5 days	5 days	5 days	5 days	na	na	na
KCl	na	na	na	4 weeks	na	na	4 weeks	na	na	na	na	na	na	na	na
NH4OAc	na	na	na	na	na	na	na	na	5 days	5 days	5 days	5 days	na	na	na
2% HNO3	na	6 months	na	na	na	na	na	na	6 months	6 months	6 months	6 months	na	na	na
Double acid	na	<5 days	na	na	na	na	na	na	<5 days	<5 days	<5 days	<5 days	na	na	na
Bray	na	na	<5 days	na	na	na	na	na	na	na	na	na	na	na	na

*Note: acidified samples for TP have a 28 day shelf life

** Some samples can be frozen and run at a later date. Check with lab manager.

1. Environmental Protection Agency, Code of Federal Regulations, 40CFR 136.3,(revised 2002).

Rain gauge bottles receive the addition of 1ml of PMA solution as a preservative. See Procedures for Chemical Analysis, page 4 for further information.

Appendix C - Required Containers, Preservation Techniques, and Holding Times^I

Shaded gray areas indicate currently tested at Coweeta

Parameter No./name time ⁴	Container ¹	Preservation ^{2,3}	Maximum holding
Table IA—Bacteria Tests:			
1–4 Coliform, fecal and total	P, G	Cool, 4C, 0.008% Na ₂ S ₂ O ₃ ⁵ ...	6 hours.
5 Fecal streptococci	P, G	Cool, 4C, 0.008% Na ₂ S ₂ O ₃ ⁵ ...	6 hours.
Table IA—Aquatic Toxicity Tests:			
6–10 Toxicity, acute and chronic	P, G	Cool, 4 °C ¹⁶	36 hours.
Table IB—Inorganic Tests:			
1. Acidity	P, G	Cool, 4°C	14 days.
2. Alkalinity	P, G	do	Do.
4. Ammonia	P, G	Cool, 4°C, H ₂ SO ₄ to pH<2 ..	28 days.
9. Biochemical oxygen demand	P, G	Cool, 4°C	48 hours.
10. Boron	P, PFTE, or Quartz.	HNO ₃ TO pH<2	6 months.
11. Bromide	P, G	None required	28 days.
14. Biochemical oxygen demand, carbonaceous ..	P, G	Cool, 4°C	48 hours.
15. Chemical oxygen demand	P, G	Cool, 4°C, H ₂ SO ₄ to pH<2 ...	28 days.
16. Chloride	P, G	None required	Do.
17. Chlorine, total residual	P, G	do	Analyze immediately.
21. Color	P, G	Cool, 4°C	48 hours.
23–24. Cyanide, total and amenable to chlorination.	P, G	Cool, 4°C, NaOH to pH>12, 0.6g ascorbic acid	14 days ⁶
25. Fluoride	P	None required	28 days.
27. Hardness	P, G	HNO ₃ to pH<2, H ₂ SO ₄ to pH<2 ..	6 months.
28. Hydrogen ion (pH)	P, G	None required	Analyze immediately.
31, 43. Kjeldahl and organic nitrogen	P, G	Cool, 4°C, H ₂ SO ₄ to pH<2 ..	28 days.
Metals: ⁷			
18. Chromium VI	P, G	Cool, 4°C	24 hours.
35. Mercury	P, G	HNO ₃ to pH<2	28 days.
3, 5–8, 12, 13, 19, 20, 22, 26, 29, 30, 32–34, 36, 37, 45, 47, 51, 52, 58–60, 62, 63, 70–72, 74,			
75. Metals, except boron, chromium VI and mercury.	P, G	do	6 months.
38. Nitrate	P, G	Cool, 4°C	48 hours.
39. Nitrate-nitrite	P, G	Cool, 4°C, H ₂ SO ₄ to pH<2	28 days.
40. Nitrite	P, G	Cool, 4°C	48 hours.
41. Oil and grease	G	Cool to 4°C, HCl or H ₂ SO ₄ to pH<2.....	28 days.
42. Organic Carbon	P, G	Cool to 4 °C HCl or H ₂ SO ₄ or H ₃ PO ₄ , to pH<2...28 days.	
44. Orthophosphate	P, G	Filter immediately, Cool, 4°C.....	48 hours.
46. Oxygen, Dissolved Probe	G Bottle and top.	None required	Analyze immediately.
47. Winkler	do	Fix on site and store in dark.....	8 hours.
48. Phenols	G only	Cool, 4°C, H ₂ SO ₄ to pH<2	28 days.
49. Phosphorus (elemental)	G	Cool, 4°C	48 hours.
50. Phosphorus, total	P, G	Cool, 4°C, H ₂ SO ₄ to pH<2	28 days.
53. Residue, total	P, G	Cool, 4°C	7 days.
54. Residue, Filterable	P, G	do	7 days.
55. Residue, Nonfilterable (TSS)	P, G	do	7 days.
56. Residue, Settleable	G	do	48 hours.
57. Residue, volatile	P, G	do	7 days.
61. Silica	P, PFTE, or Quartz.	Cool, 4 °C	28 days.
64. Specific conductance	P, G	do	Do.
65. Sulfate	P, G	do	Do.
66. Sulfide	P, G... ..	Cool, 4°C add zinc acetate plus sodium hydroxide to pH>9	7 days.
67. Sulfite	P, G	None required	Analyze

immediately.

68. Surfactants	P, G	Cool, 4°C	48 hours.
69. Temperature	P, G	None required	Analyze.
73. Turbidity	P, G	Cool, 4°C	48 hours.

Table IC—Organic Tests⁸

13, 18–20, 22, 24–28, 34–37, 39–43, 45–47, 56,

76, 104, 105, 108–111, 113. Purgeable Halocarbons. G, Teflonlined septum. Cool, 4 °C, 0.008% Na₂S₂O₃⁵. 14 days.

6, 57, 106. Purgeable aromatic hydrocarbonsdo Cool, 4 °C, 0.008% Na₂S₂O₃⁵ HCl to pH2⁹. Do.

3, 4. Acrolein and acrylonitriledo Cool, 4 °C, 0.008% Na₂S₂O₃⁵ adjust pH to 4–5¹⁰ Do.

23, 30, 44, 49, 53, 77, 80, 81, 98, 100, 112. Phenols¹¹. G, Teflonlinedcap..Cool, 4 °C, 0.008% Na₂S₂O₃⁵ 7 days until extraction; 40 days after extraction.

7, 38. Benzidines¹¹dodo 7 days until extraction.¹³

14, 17, 48, 50–52. Phthalate esters¹¹do Cool, 4 °C 7 days until extraction; 40 days after extraction.

82–84. Nitrosamines^{11,14}do Cool, 4 °C, 0.008% Na₂S₂O₃⁵ store in dark. Do.

88–94. PCBs¹¹do Cool, 4 °C Do.

54, 55, 75, 79. Nitroaromatics and isophorone¹¹do Cool, 4 °C, 0.008% Na₂S₂O₃⁵ store in dark. Do.

1, 2, 5, 8–12, 32, 33, 58, 59, 74, 78, 99, 101. Polynuclear aromatic hydrocarbons¹¹.....dodo Do.

15, 16, 21, 31, 87. Haloethers¹¹do Cool, 4 °C, 0.008% Na₂S₂O₃⁵ Do.

29, 35–37, 63–65, 73, 107. Chlorinated hydrocarbons¹¹.....do Cool, 4 °C Do.

60–62, 66–72, 85, 86, 95–97, 102, 103. CDDs/CDFs¹¹aqueous: field and lab preservation. .. G Cool, 0–4 °C, pH<9, 0.008% Na₂S₂O₃. 1 year.

Solids, mixed phase, and tissue: field preservation.....do Cool, <4 °C 7 days.

Solids, mixed phase, and tissue: lab preservationdo Freeze, <¥10 °C 1 year.

Table ID—Pesticides Tests:

1–70. Pesticides¹¹do Cool, 4°C, pH 5–9¹⁵ Do.

Table IE—Radiological Tests:

1–5. Alpha, beta and radium P, G HNO₃ to pH<2 6 months.

Appendix C-- Notes

1. Polyethylene (P) or glass (G). For microbiology, plastic sample containers must be made of sterilizable materials (polypropylene or other autoclavable plastic).

2. Sample preservation should be performed immediately upon sample collection. For composite chemical samples each aliquot should be preserved at the time of collection. When use of an automated sampler makes it impossible to preserve each aliquot, then chemical samples may be preserved by maintaining at 4°C until compositing and sample splitting is completed.

3. When any sample is to be shipped by common carrier or sent through the United States Mails, it must comply with the Department of Transportation Hazardous Materials Regulations (49 CFR part 172). The person offering such material for transportation is responsible for ensuring such compliance. For the preservation requirements of Table II, the Office of Hazardous Materials, Materials Transportation Bureau, Department of Transportation has determined that the Hazardous Materials Regulations do not apply to the following materials: Hydrochloric acid (HCl) in water solutions at concentrations of 0.04% by weight or less (pH about 1.96 or greater); Nitric acid (HNO₃) in water solutions at concentrations of 0.15% by weight or less (pH about 1.62 or greater); Sulfuric acid (H₂SO₄) in water solutions at concentrations of 0.35% by weight or less (pH about 1.15 or greater); and Sodium hydroxide (NaOH) in water solutions at concentrations of 0.080% by weight or less (pH about 12.30 or less).

4. Samples should be analyzed as soon as possible after collection. The times listed are the maximum times that samples may be held before analysis and still be considered valid. Samples may be held for longer periods only if the permittee, or monitoring laboratory, has data on file to show that for the specific types of samples under study, the analytes are stable for the longer time, and has received a variance from the Regional Administrator under § 136.3(e). Some samples may not be stable for the maximum time period given in the table. A permittee, or monitoring laboratory, is obligated to hold the sample for a shorter time if knowledge exists to show that this is necessary to maintain sample stability. See § 136.3(e) for details. The term “analyze immediately” usually means within 15 minutes or less of sample collection.

5. Should only be used in the presence of residual chlorine.

6. Maximum holding time is 24 hours when sulfide is present. Optionally all samples may be tested with lead acetate paper before pH adjustments in order to determine if sulfide is present. If sulfide is present, it can be

removed by the addition of cadmium nitrate powder until a negative spot test is obtained. The sample is filtered and then NaOH is added to pH 12.

7. Samples should be filtered immediately on-site before adding preservative for dissolved metals.

8. Guidance applies to samples to be analyzed by GC, LC, or GC/MS for specific compounds.

9. Sample receiving no pH adjustment must be analyzed within seven days of sampling.

10. The pH adjustment is not required if acrolein will not be measured. Samples for acrolein receiving no pH adjustment must be analyzed within 3 days of sampling.

11. When the extractable analytes of concern fall within a single chemical category, the specified preservative and maximum holding times should be observed for optimum safeguard of sample integrity. When the analytes of concern fall within two or more chemical categories, the sample may be preserved by cooling to 4°C, reducing residual chlorine with 0.008% sodium thiosulfate, storing in the dark, and adjusting the pH to 6–9; samples preserved in this manner may be held for seven days before extraction and for forty days after extraction. Exceptions to this optional preservation and holding time procedure are noted in footnote 5 (re the requirement for thiosulfate reduction of residual chlorine), and footnotes 12, 13 (re the analysis of benzidine).

12. If 1,2-diphenylhydrazine is likely to be present, adjust the pH of the sample to 4.0±0.2 to prevent rearrangement to benzidine.

13. Extracts may be stored up to 7 days before analysis if storage is conducted under an inert (oxidant-free) atmosphere.

14. For the analysis of diphenylnitrosamine, add 0.008% Na₂S₂O₃ and adjust pH to 7–10 with NaOH within 24 hours of sampling.

15. The pH adjustment may be performed upon receipt at the laboratory and may be omitted if the samples are extracted within 72 hours of collection. For the analysis of aldrin, add 0.008% Na₂S₂O₃.

16. Sufficient ice should be placed with the samples in the shipping container to ensure that ice is still present when the samples arrive at the laboratory. However, even if ice is present when the samples arrive, it is necessary to immediately measure the temperature of the samples and confirm that the 4°C temperature maximum has not been exceeded. In the isolated cases where it can be documented that this holding temperature can not be met, the permittee can be given the option of on-site testing or can request a variance. The request for a variance should include supportive data which show that the toxicity of the effluent samples is not reduced because of the increased holding temperature.

Appendix D

Allowable Percent Error by Vendor of Certified QC Standards

<u>Method</u>	<u>Analyte</u>	<u>Allowable ± %Error¹</u>	<u>Found %Absolute Accuracy²</u>
ICP,AA	K	15	10.6, 2.99
	Na	15	8.05, 2.28
	Ca	14	11.6, 2.26
	Mg	14	10.6, 2.24
ICP	P	19	4.20
	Al	21	13.6
Auto Analyzer	NH ₄	16	6.67
	NO ₃	10	3.05
	SiO ₃	15	5.74
	PO ₄	14	5.16
Ion Chromatograph	Cl	10	5.55
	NO ₃	10	3.48
	PO ₄	10	6.18
	SO ₄	10	4.12
	Br	10	5.75
Shimadzu	DOC	17	5.06
	TN	23	3.23
pH meter	pH	2.4	.032

1. The vendor certifying the quality control sample has a certified value and a range associated with that value. This range is the allowable percent error. The ranges are based on actual historical data collected by the vendor. The range reflects any bias in the method used to establish the limits and closely approximate a 95% confidence interval of the performance that experienced laboratories should achieve using acceptable environmental methods.
2. %Absolute Accuracy was the result of analysis in 2013 for Quarterly QC values for Coweeta.

Appendix E – Method Detection Limits by Sample Type

Water

Analyte	Lab Designation	Method	Instrument	Instrument in Operation	Units of reported values	2013 Matrix =DI
Ammonium-Nitrogen	NH4-N	automated Phenate method	Astoria 2 Autoanalyzer, Astoria-Pacific, Clackamas, Oregon	11/14/2006	mg/L	0.002
Nitrate-Nitrogen	NO3-N	reduction using a cadmium coil	Astoria 2 Autoanalyzer, Astoria-Pacific, Astoria, Oregon	11/14/2006	mg/L	0.002
Potassium	K	Flame - absorption	Perkin Elmer Analyst300 Atomic Absorption Spectrometer, Perkin Elmer, Waltham, MA	June 1999	mg/L	0.004
Sodium	Na	Flame - absorption	Perkin Elmer Analyst300 Atomic Absorption Spectrometer, Perkin Elmer, Waltham, MA	June 1999	mg/L	0.018
Calcium	Ca	Flame - absorption	Perkin Elmer Analyst300 Atomic Absorption Spectrometer, Perkin Elmer, Waltham, MA	June 1999	mg/L	0.018
Magnesium	Mg	Flame - absorption	Perkin Elmer Analyst300 Atomic Absorption Spectrometer, Perkin Elmer, Waltham, MA	June 1999	mg/L	0.002
Potassium	K	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.018
Sodium	Na	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.012
Calcium	Ca	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.025
Magnesium	Mg	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.008
Aluminum	Al	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.009
phosphorous	P	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.008
Sulfur	S	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.010

Chloride	Cl	Micro-membrane Suppressed Ion Chromatography , using an AS 18 column	Dionex 2500 Ion Chromatograph, from Dionex, Sunnyvale, CA	Sept 2004	mg/L	0.011
Bromide	Br	Micro-membrane Suppressed Ion Chromatography , using an AS 18 column	Dionex 2500 Ion Chromatograph, from Dionex, Sunnyvale, CA	Sept 2004	mg/L	0.003
Nitrate-Nitrogen	NO3-N	Micro-membrane Suppressed Ion Chromatography , using an AS 18 column	Dionex 2500 Ion Chromatograph, from Dionex, Sunnyvale, CA	Sept 2004	mg/L	0.006
ortho Phosphate (Orthophosphate is sometimes referred to as "reactive phosphorus.")	PO4	Micro-membrane Suppressed Ion Chromatography , using an AS 18 column	Dionex 2500 Ion Chromatograph, from Dionex, Sunnyvale, CA	Sept 2004	mg/L	0.008
Sulfate	SO4	Micro-membrane Suppressed Ion Chromatography , using an AS 18 column	Dionex 2500 Ion Chromatograph, from Dionex, Sunnyvale, CA	Sept 2004	mg/L	0.003
Dissolved organic carbon	DOC	catalytically-aided platinum 680°C combustion technique for sample oxidation	Shimadzu DOC-VCPH TN analyzer, Shimadzu Scientific Instruments, Columbus, MD	9/20/2005	mg/L	0.011
Total dissolved nitrogen	TN	luminescence	Shimadzu DOC-VCPH TN analyzer, Shimadzu Scientific Instruments, Columbus, MD	9/20/2005	mg/L	0.038
Hydrogen ion activity in in water	pH for Water	Standard Methods using a Broadley James pH combo probe, Broadley James Corp, Irvine CA	Orion Research Digital pH/millivolt 611 meter, Thermo Fisher Scientific, Waltham, MA	1985	pH units	0.010

Continued

SOIL

Analyte	Lab Designation	Method	Instrument	Instrument in Operation	Units of reported values	2013 1N NH4Cl high	2013 Bray	2013 DA	2013 1N NH4OAc	2013 1N KCl
Ammonium-Nitrogen	NH4-N	automated Phenate method	Astoria 2 Autoanalyzer, Astoria-Pacific, Clackamas, Oregon	11/14/2006	mg/L					0.005
Nitrate-Nitrogen	NO3-N	reduction using a cadmium coil	Astoria 2 Autoanalyzer, Astoria-Pacific, Astoria, Oregon	11/14/2006	mg/L					0.003
Soluble reactive phosphorus	P	Ammonium Molybdate / Antimony Potassium Tartrate reaction and reduction with Ascorbic Acid	Astoria 2 Autoanalyzer, Astoria-Pacific, Astoria, Oregon	11/14/2006	mg/L		0.014			
Potassium	K	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.134			0.043	
Sodium	Na	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.085				
Calcium	Ca	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.070			0.017	
Magnesium	Mg	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.058			0.065	
Aluminum	Al	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.235				
phosphorous	P	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L		0.014	0.005		
Analyte	Lab Designation	Method	Instrument	Instrument in Operation	Units of reported values	2013 %Soil Std = Thermo				
carbon	C	dumas method	Flash EA 1112 NC analyzer, CE Elantech, Lakewood, NJ	9/15/05	%	0.153				
nitrogen	N	dumas method	Flash EA 1112 NC analyzer, CE Elantech, Lakewood, NJ	9/15/05	%	0.019				
Hydrogen ion activity in in water	pH for soil	Standard Methods using an Orion 9165BN combination pH electrode	Thermoscientific Orion 3 star pH benchtop	August 2009	pH units		0.016			

Continued

Plant

Analyte	Lab Designation	Method	Instrument	Instrument in Operation	Units of reported values	2013 Matrix= 2%HNO3
Potassium	K	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.106
Sodium	Na	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.088
Calcium	Ca	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.102
Magnesium	Mg	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.084
Aluminum	Al	Optical Emission	Thermo Fisher iCAP 6300, Madison WI	November 30,2012	mg/L	0.087
Analyte	Lab Designation	Method	Instrument	Instrument in Operation	Units of reported values	2013 plant tissue Peach Leaves %
carbon	C	dumas method	Flash EA 1112 NC analyzer, CE Elantech, Lakewood, NJ	9/15/05	%	1.257
nitrogen	N	dumas method	Flash EA 1112 NC analyzer, CE Elantech, Lakewood, NJ	9/15/05	%	0.101